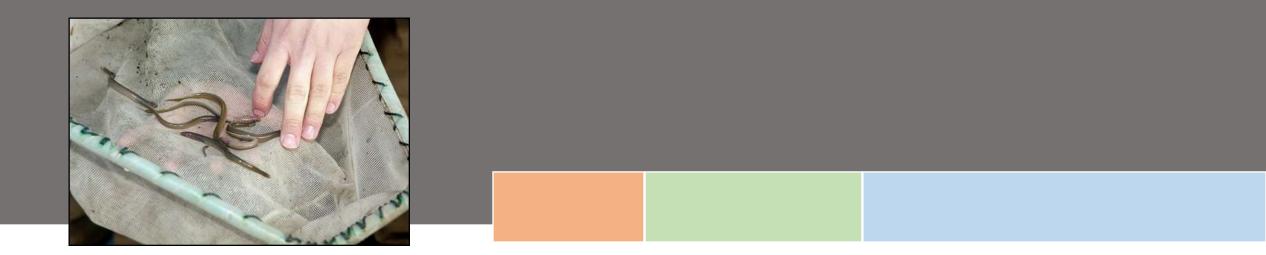
The Shocking Story of Larval Lamprey



Joe Krieter, M.S. - Senior Aquatic Ecologist Bill Allen – Electronics Specialist Hart Crowser, Inc. – Portland, Oregon



Project Sponsor and Collaborative Partners

Project Sponsor: Michelle Hollis, M.S.

Port of Portland, Environmental Ops., Portland, OR

eDNA Lab: Taal Levi, Ph.D. and Jennifer Allen

Oregon State University, Quantitative Wildlife Ecology and Conservation Lab, Corvallis, OR

Lamprey Hatchery: Ralph Lampman

Yakama Nation FRMP, Pacific Lamprey Project, Prosser, WA

+ USACE, USFWS, ODFW, and Battelle Labs





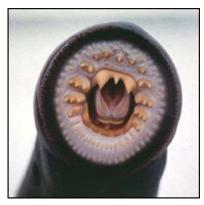




What are Lamprey?

- Class Agnatha, the jawless fishes
- Family Petromyzonidae (stone suckers)
- Round, elongated body
- No paired fins, scales, jaws, or bones (only cartilage)
- Round sucker-like mouth for clinging and sucking
- Often mistaken for eels (however, eels have jaws)
- Lack swim bladders and must swim constantly or hold onto objects
- Adult lamprey can grow to up to three feet in length
- Feed on bodily fluids of fish and marine mammals

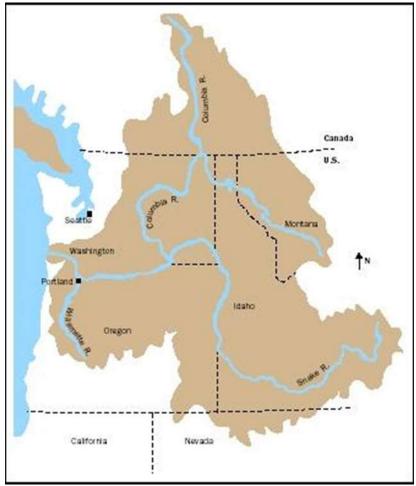






Lamprey Species of the Columbia and Willamette River Basin

- **Pacific Lamprey** (*Entosphenus tridentatus,* formerly *Lampetra tridentata,* "three-toothed stone sucker")
- Western Brook Lamprey (Lampetra richardsoni)
- River Lamprey (Lampetra ayresi)





Why the interest in Lamprey?

- They are just way cool (and cute?)!
 - Class Agnatha, or jawless fishes, the most primitive of all living vertebrates
 - Oldest fish alive today with a fossil record dating back 500 million years
- Play a vital role in the ecosystem as food for mammals, fish and birds, and for nutrient cycling and storage
- Indigenous peoples harvested lampreys for subsistence, religious, medicinal, spiritual and cultural purposes for many generations







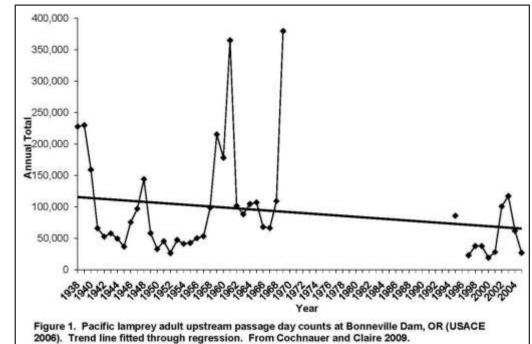
River Monsters – "Vampires of the Deep"

Jeremey Wade -Discovery's Animal Planet 2013



Federal and State Status

- Not currently "listed" as threatened or endangered
- Petitioned in January 2003
 - Pacific lamprey, western brook lamprey, river lamprey, and Kern brook lamprey
 - Oregon, Washington, Idaho, and California
 - Based on a decline in abundance and distribution throughout their range
- In December 2004 the USFWS determined that there was not substantial scientific or commercial information to warrant listing



Lamprey Species	Status		
	Federal	OR	WA
Pacific lamprey	SOC	SV	Monitor
River lamprey	SOC	None	-
Western brook lamprey	None	SV	Monitor



USFWS 2010 Best Management Practices

Reasons for Decline:

 lack of passage: dewatering and reduced flows: poisoning; poor water quality; dredging (channel maintenance and mining); stream and floodplain degradation (channelization, loss of side channel habitat, scouring); ocean conditions (loss of prey, increase in predators); predation by nonnative fish species

Conservation Measures:

- provide lamprey passage
- protect ammocoete habitat
- restore stream channel complexity







Adults live in ocean 1-3 years and feed on host fish



Adults migrate to freshwater and reside there about a year



Adults spawn in gravel nest then die

Lamprey Life Cycle

Pacific



Adults develop teeth on sucking disk for parasitic feeding





Larvae transform to juveniles (macropthalmia) and migrate to the ocean



Ammocoetes live in silt/sand substrates and filter feed for 3 - 7 years



Eggs hatch into larvae (ammocoetes) and drift downstream to slow velocity area

Streif, B. 2009. American Fisheries Society Symposium 73. Published by the American Fisheries Society. ISBN: 978-1-934874-13-4



Pacific Lamprey Larva (Ammocoetes)





Permit Conditions - Abbreviated

Terminal-Wide Berth Maintenance Program USACE Permit Special Condition #7 - Develop a plan addressing Pacific lamprey that may be impacted by berth maintenance activities



Challenges and Solutions

- Many unknowns regarding the presence of ammocoetes
 - Low density, patchy distribution, very little habitat preference information
- Very challenging sampling conditions and restrictions
 - 45+ ft. water depths, limited visibility
 - Very small and burrowed organisms that are highly mobile
 - Water temperature restrictions on electrofishing (64.4°F)
- Few successful sampling methods
 - USFWS electrified suction dredge
 - PNNL deep water electrofishing platform

Mueller, R.P., E.V. Arntzen, M. Nabelek, B.L. Miller, K. Klett & R.A. Harnish. 2012. Laboratory testing of a modified electroshocking system designed for deepwater juvenile lamprey sampling. Transactions of the American Fisheries Society 141(3):841-845





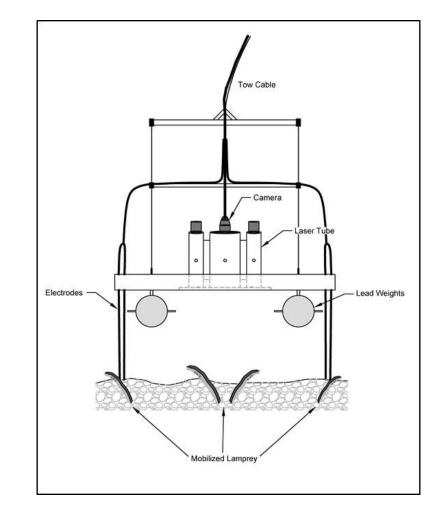
Larval Lamprey Electrofishing System (LLES)

Operator console

- Modified ETS electrofishing unit
- Camera and LED controls
- Dual color video monitors
- DVR

Sampling Sled

- Two hydraulic sounding weights
- Dual electrofishing anodes / cathodes
- Low-light color drop camera
- Paired scaling lasers
- External lighting equipment
- Custom welded frame and sheathed cable bundle





LLES Advantages and Disadvantages

<u>Advantages</u>

- Builds upon prior accepted studies
- Relatively efficient sampling
- Small crew

Disadvantages

- Requires clear water conditions
- Cannot distinguish ammocoetes by species





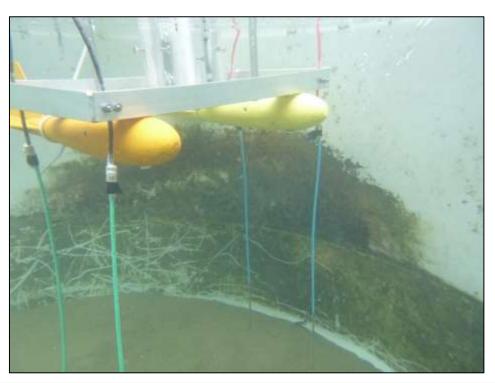




LLES Testing – Prosser Fish Hatchery

- Known density, species and origin (10 PL / m²)
- Three trials conducted
- Resulting in 94% to 57% accuracy; similar to prior studies











LLES Sampling

- Three shipping terminals, one reference site;
- 12 transects, 400 sampling stations, 50 ft. x 50 ft. grid;
- 30-60 seconds @150 VDC, 4 Hz, 25 percent duty cycle and a 3:1 burst pulse rate;
- 3-7 minutes for maneuvering between stations;
- Approx. 3 ft² sampling area, 18 inches off the bottom.

















Sediment Grab Sampling

- Collected for eDNA analysis for a subset (10%) of sites
- Petite ponar dredge sampler
- Sterile techniques
- One 50 mL sample and two replicates
- Delivered frozen to OSU for extraction and ddPCR







🐼 OSU 🌌

eDNA

 "Genetic material obtained directly from environmental samples (soil, sediment, water, etc.) without any obvious signs of biological source material"

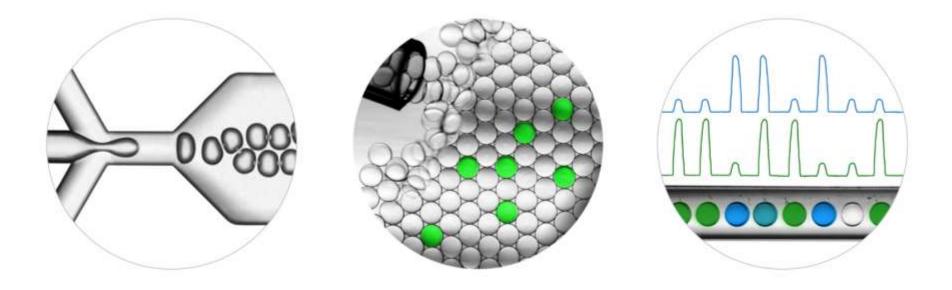
From:

- Advantages
 - Non-invasive
 - Highly sensitive
 - Short degradation time
 - Cost-efficient
- Disadvantages
 - No info. about DNA source (life stage, condition, etc.)
 - DNA can be detected from sources other than target

P.H. Thomsen and E. Willersley. 2015. Environmental DNA
– An Emerging Tool in Conservation for Monitoring Past and
Present Biodiversity. Biological Conservation. Vol 183. pp. 4 – 18.

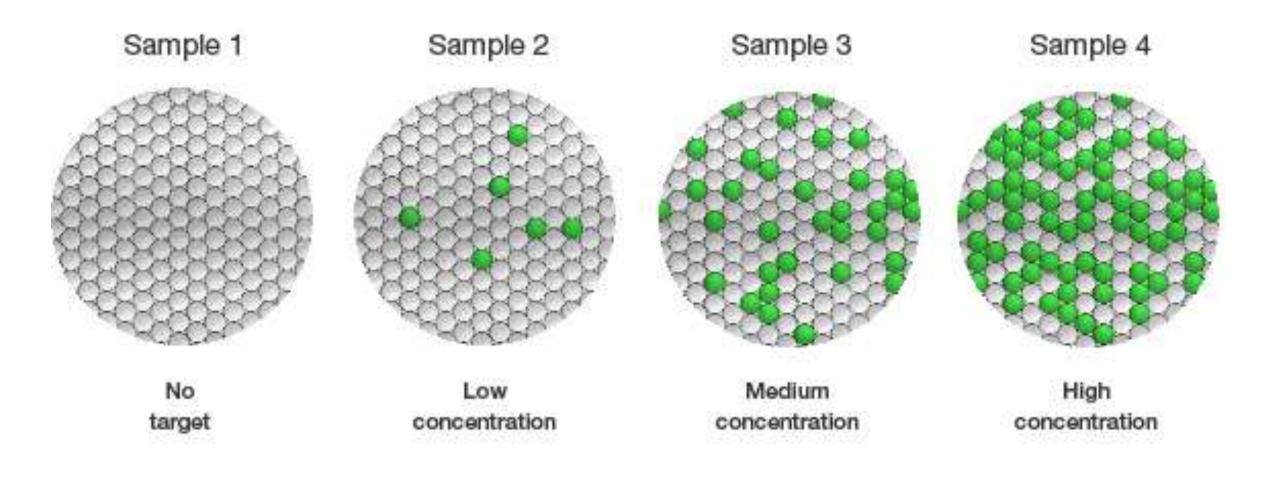


Absolute quantification with Droplet digital PCR



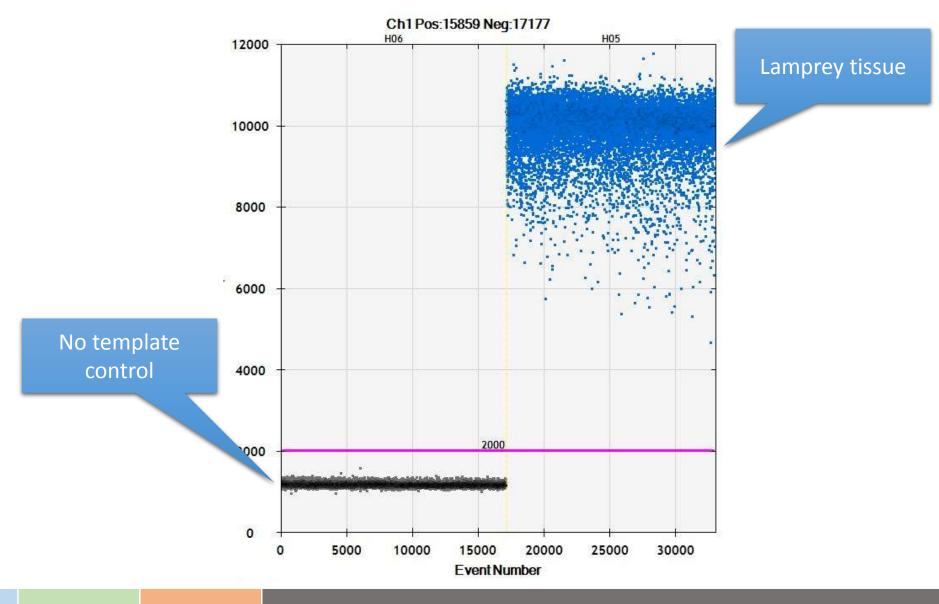


Absolute Quantification





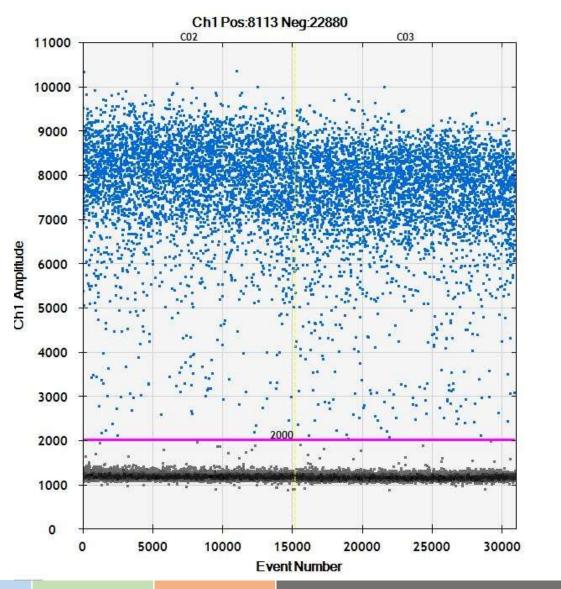
Positive and Negative Controls

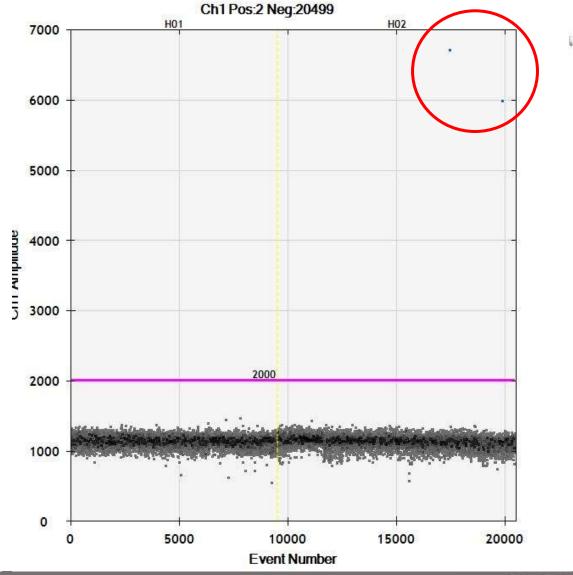




Abundant

Rare





🗞 OSU 🖅

Preliminary Results – Nearly There...

- Successful LLES testing results and overcoming water clarity issues upon initial deployment.
- Successful eDNA data collection, extraction and analytical techniques developed by OSU.
- Surveys and analyses incomplete at this time, but no LLES lamprey detections yet at POP berthing areas.
- Requested sampling extension from ODFW.
- Anticipate first year report in early 2018.



Thank You!

Please send us your questions and comments:

Sampling Questions:

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